

Blend of chitosan acetate salt with poly(*N*-vinyl-2-pyrrolidone): Interaction between chain-chain

Shuguang Cao, Yanqiao Shi, Guanwen Chen*

Institute of Chemistry, Chinese Academy of Sciences, Beijing 100080, P.R. China

Received: 1 June 1998/Revised version: 1 October 1998/Accepted: 13 October 1998

SUMMARY: Clear blends of chitosan acetate salt with poly (*N*-vinyl-2-pyrrolidone) (PVP) made from aqueous solutions look to be miscible on appearance. IR spectra investigated the carbonyl-hydroxyl hydrogen bonding between chain-chain in the blends and thus evidenced the compatibility of two polymers on a molecular level. The IR spectra, DSC also explored the interaction changing during the temperature increasing and indicated that a significant conformational change took place.

Introduction

Chitin is one of the most abundant natural polymers being second only to cellulose in the amount produced annually by biosynthesis¹. It occurs in animals, particularly in crustacea, molluscs and insects, where it is an important constituent of the exoskeleton, and in certain fungi where it is the principal fibrillar polymer in the cell wall.

Chitosan is a product obtained by alkaline deacetylation of chitin and has good properties for forming membrane. In dehydration of solvents, e.g. ethanol, pyridine, the pervaporation performances were fully investigated and the results showed it was an excellent membrane^{2,3}. But it has little permeability for MeOH/MTBE mixture, probably due to its rigid chain and close packing with presence of several different types of intermolecular and intramolecular hydrogen bonding in polymer matrix, and thus up to now there is no any report on this. But with poly (*N*-vinyl-2-pyrrolidone) (PVP) blending with chitosan, the obtained polyblend membrane showed good pervaporation permeation for methanol(MeOH)/methyl tert-butyl ether(MTBE) mixture⁴. The possible explanation should contribute to the structure characteristics of polyblends, particularly the interaction between chain-chain of two amorphous polymers^{5,6}.

A number of papers reported the miscibility behavior in blends composed of polymers with dissimilar chemical structures, which provide specific interactions between chains. Miscibility of blends containing donor and acceptor polymer like those containing acid and/or basic units has been reported. It has demonstrated that poly (*N*-vinyl-2-pyrrolidone) interacts strongly with polymers of a wide variety of polymers, e.g. poly(vinylidene fluoride)⁷, polyvinylalcohol⁸, poly(2,6-dimethyl-1,4-phenylene oxide)⁹, poly (mono-*n*-alkylitaconates)¹⁰, poly (*N*-1-alkylitaconamic acids)¹¹, cellulose¹² etc. However, to our knowledge, no works dealing with blends containing chitosan acetate salt and poly (*N*-vinyl-2-pyrrolidone) have been reported.

Our earlier interest was the membrane pervaporation properties, but the membrane showed good permeability for MeOH/MTBE mixture, particularly compared with chitosan membrane. For this reason, the structure characteristics of polyblends of chitosan acetate salt and poly (*N*-vinyl-2-pyrrolidone), particularly the hydrogen-bonding interaction between chain-chain in polyblends, were investigated by IR, DSC.

Experimental

Materials

* Corresponding author

Chitosan with a degree of deacetylation of 78% and an average molecular weight of $3-4 \times 10^5$ as a membrane material was supplied by Koyo Chemical Co., Japan. Poly (*N*-vinyl-2-pyrrolidone) (PVP) with an average molecular weight of 4×10^5 was from Aldrich Co.. The other reagents and solvents were analytical-grade from commercial sources and used without further purification.

Membrane preparation

The solution of poly (*N*-vinyl-2-pyrrolidone) (1.5wt.%) and chitosan solution (1.5wt%) in 1.0wt% aqueous acetic acid were mixed in different ratios. The casting solutions of both with different composition were poured on glass plate and evaporated at room temperature. The thickness of blending membrane is about $20 \mu\text{m}$. All the samples were dried under reduced pressure in a vacuum oven at 40°C for two days before tested.

FTIR measurements

Infrared spectra of pure polymers and blends were recorded on Bruker Fourier Transform Infrared Vector 22 spectrophotometer. The spectra were recorded with a resolution of 2 cm^{-1} .

DSC Measurement

DSC curves were obtained using a Perkin Elemer DSC 7 thermal analysis system with a heating rate $20^\circ\text{C}/\text{min}$ under the nitrogen atmosphere. The samples were dried under reduced pressure in a vacuum oven at 30°C for two days.

Results and discussion

Chitosan is practically insoluble in water and organic solvents due to its micelle structure, the close packing and the presence of several different types of hydrogen bonds. Chitosan gives water-soluble salts with acetic acid (Fig.1) and thus blends with PVP to form clear films. Their transparency is dependent on the polyblend composition.

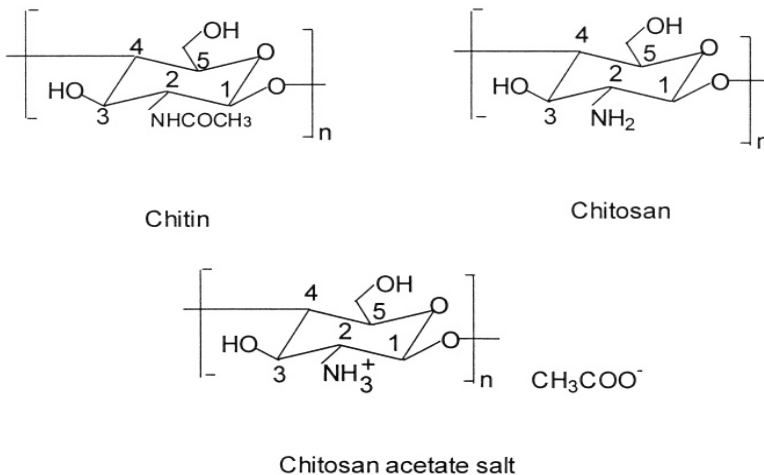


Fig.1 The chemical structure of chitin, chitosan and chitosan acetate salt

The infrared spectra of pure PVP and chitosan have been obtained^{12,13}. The main characteristic bands are the following:

3480 cm^{-1} C₆-OH stretching band of chitosan

3447 cm^{-1} C₃-OH stretching band of chitosan

3375 cm^{-1} hydrogen bonding of amide

3264 cm^{-1} N-H stretching band

1681 cm^{-1} C=O stretching of PVP

1652 cm^{-1} amide I band of chitosan

1558 cm^{-1} amide II band of chitosan, overlapping with NH₃⁺ band(1500 cm^{-1})

The good identification of these peaks allows us to investigate the influence of chain-chain interaction between the polyblend on the basis of band positions and relative intensities.

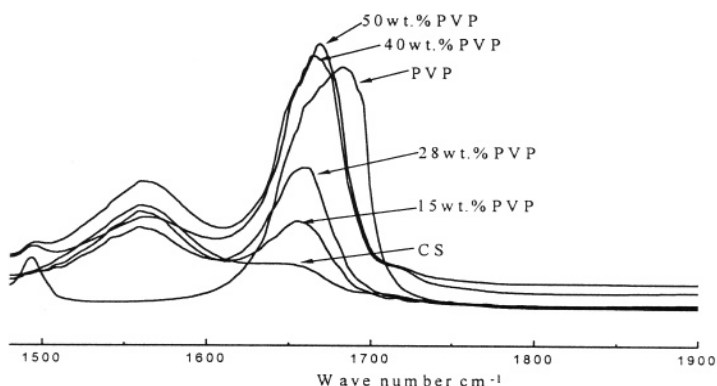


Fig.2 IR spectra of chitosan/PVP blends in region of 1300-1900 cm^{-1}

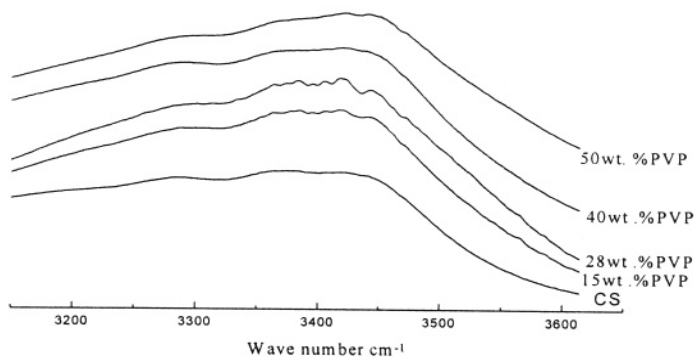


Fig.3 IR spectra of chitosan/PVP blends in region of 3150-3600 cm^{-1}

The chain-chain interaction between two polymers can be deduced from the shifts of the vibration frequencies of the group involved in the hydrogen bonding, e.g., the

frequencies of the PVP carbonyl (Fig.2) and those of the O-H bands of chitosan (Fig.3). When the PVP content increases, the carbonyl band shifts towards lower frequencies and finally overlaps with the amide I band, indicating that carbonyl of PVP takes part in hydrogen bonding with chitosan. With increasing the PVP content, strong NH_3^+ band overlapping with amide II band at 1558cm^{-1} exists in polyblends and has no obvious shift, indicating that the NH_3^+ does not participate the intermolecular bonding between chitosan and PVP. So the carbonyl of PVP should form hydrogen bonding with hydroxyl of chitosan. This result also proves that there is miscibility at a molecular level between two polymers, at least in some regions. The C-O stretching band of chitosan near 1100cm^{-1} shifts steadily towards higher frequencies when the PVP content increasing (Fig.4). The possible explanation should be: in the pure chitosan film, the C-O-H groups involve in different types of self-associations; the C-O stretching frequency is lower than the frequency of a free C-O-H group. With the PVP incorporating into chitosan, many hydroxyl groups are involved in hydroxyl-carbonyl hydrogen bonding at the expense of hydroxyl-hydroxyl hydrogen bonding. As a result many oxygen atoms of C-OH are freed from hydrogen bonding. Meanwhile, a similar phenomenon may happen with the OH stretching band (Fig.3). These bands are usually assigned to the hydrogen-bonded hydroxyls. The splitting of the hydroxyl band of chitosan upon blending with PVP helps us to further insight into the interaction in the system. Since the bands in region of $3300\text{-}3500\text{cm}^{-1}$ are usually assigned to the inter- and intramolecular hydrogen bonding of hydroxyl groups, the occurrence of splitting in this region indicates that the carbonyl group of PVP take part in the hydrogen bonding with hydroxyl group and thus change the hydrogen bonding interaction results in matrix. This result correlate with pervaporation properties that the membrane performs high permeation with PVP added in chitosan to alternate the interaction in matrix. According to the splitting condition in Fig.3, it is supposed that there are existed the following forms of hydrogen bonding after blending with PVP (Fig.5): 1) hydroxyl groups associated by multiple hydrogen bonding in matrix, 2) hydroxyl groups whose proton is involved in hydrogen bonding with ca carbonyl group and an oxygen atom of another hydroxyl group, or with two carbonyl groups, 3) hydroxyl groups simply bonded to a carbonyl group.

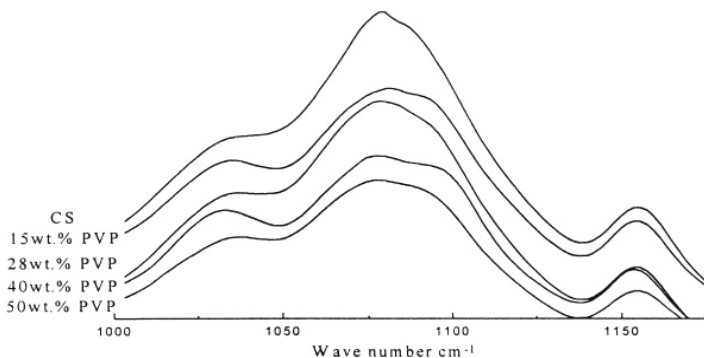


Fig.4 IR spectra of chitosan/PVP blends in region of $1000\text{-}1200\text{cm}^{-1}$

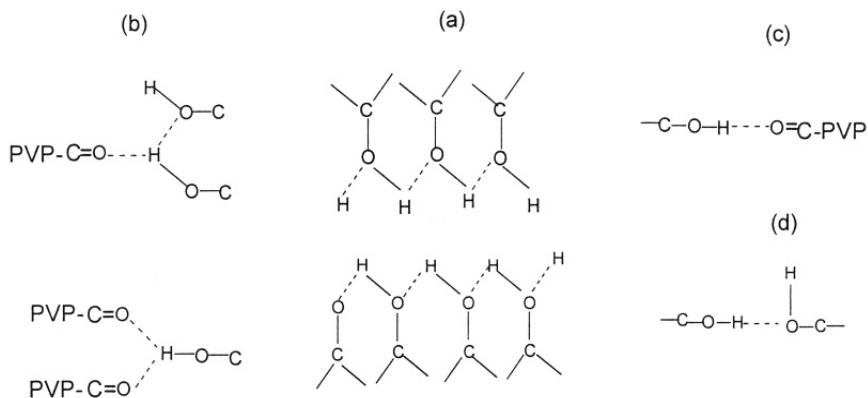


Fig.5 Different types of hydrogen bonding of hydroxyl groups of chitosan in the blends (a) Inter and intramolecular hydrogen bonding in matrix, (b) cross-interaction of hydroxyl group di-hydrogen bonded with two carbonyl groups or with one carbonyl and one oxygen atom of another hydroxyl group, (c) cross-interaction between a PVP carbonyl group and a chitosan hydroxyl group, (d) simple hydrogen bonding between two hydroxyl groups.

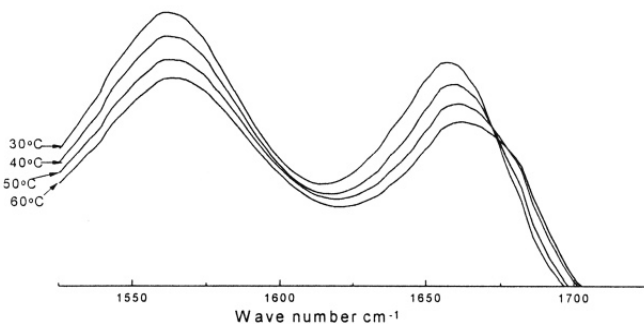


Fig.6 Influence of temperature on IR spectra of blend with 28wt.%PVP content in region of 1450-1700 cm^{-1}

Additionally, it is also found that the blending membrane is sensitive to the temperature (Fig.5). When the temperature increasing from 30 to 60 $^{\circ}\text{C}$, it is the carbonyl band of the PVP, not the amide I band of chitosan because the amide II band and the amide hydrogen bonding (3375 cm^{-1}) have no any shift, which shifts to higher

frequencies. This phenomenon indicates that the hydrogen-carbonyl hydrogen bonding is reducing with improving the chain motion. In fact, with the temperature increasing, the changing of the peaks in $3000\text{-}3600\text{cm}^{-1}$ region also confirms the above conclusion (Fig.6). The peaks at 3264cm^{-1} and 3375cm^{-1} don't obviously change, but the relatively intensities of peaks at 3447cm^{-1} and 3480cm^{-1} have some change. Generally these bands are assigned to hydroxyl groups associated by multiple hydrogen bonds with hydroxyl groups, carbonyl groups ect⁸. So this temperature dependence is indicative of bands that reflect conformational sensitivity and suggests that a significant rearrangement of bond structure may be taking place within the blending membranes. In fact, DSC results of blending membrane with 28wt.% PVP content showed that there

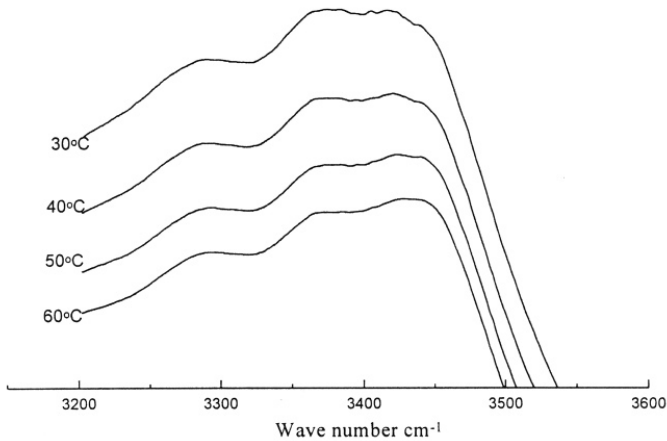


Fig.7 Influence of temperature on IR spectra of blend with 28wt.%PVP content in region of $3250\text{-}3500\text{cm}^{-1}$

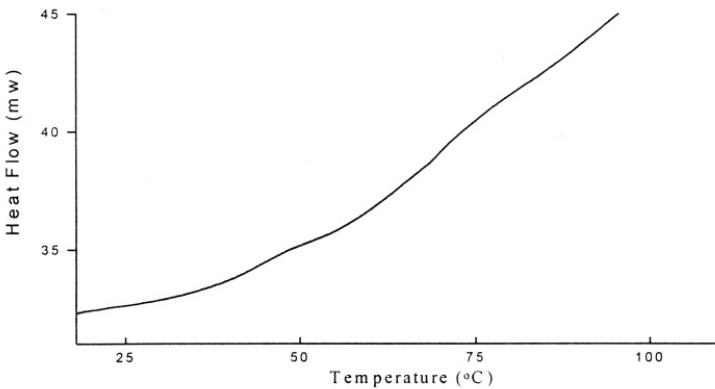


Fig.8 The DSC curves of blend membrane with 28wt.% PVP content

exited two second transition at 40.4°C and 56.2°C, respectively (Fig.7). Undoubtedly, these second transitions should contribute the chain conformational change during the temperature increasing and thus result in the interaction changing.

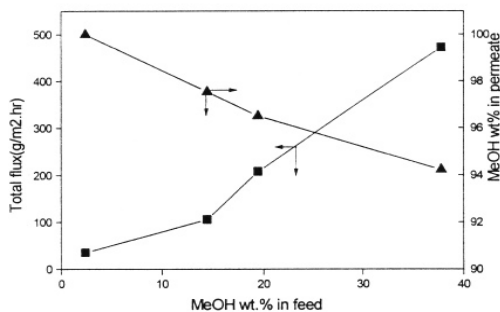


Fig.9 The pervaporation properties of blend membrane with 28wt.%PVP content

The above mentioned showed that the structure of membrane has been changed after the PVP added in matrix, particularly the hydrogen bonding forms. The blending membrane with 28wt.%PVP content was evaluated for separation of MeOH from MTBE. The pervaporation results are given in Fig.9 as function of the MeOH content in feed. It is shown that the blend membrane performed good permeability, especially compared with chitosan(almost no permeation). The possible explanation should be that the blending changed the rigid chain and close packing structure in membrane. Meanwhile, the carbonyl groups of PVP also form hydrogen bonding with MeOH in feed and thus act on sorption sites in pervaporation process. As a result, the blend membrane showed MeOH preferential permeation.

Acknowledgements This work was supported by Natural Science Foundation of China. The authors also thank Dr.Zhang Shimin for helpful discussions.

Reference:

1. M.V.Tracey(1957) Rev.Pure Chem. 7:1
2. T.Urayama, K.Takigawa, (1990) Polymer 31:668
3. B.K.Oh, C.K.Park and Y.M.Lee (1993) Polymer Bulletin 31:723
4. S.Cao, Y.Shi and G.Chen, Makromol.Chem. submitted.
5. Z.Ping, Q.Nguyen and J.Neel(1989) Makromol.Chem., 190:437
6. A.Wrzyszczynski et al (1995) Polymer Bulletin 34:493
7. M.Galin (1987) Makromol.Chem., 188:1391
8. Z.Ping, Q.Nguyen and J.Neel(1990) Makromol.Chem., 191:185
9. A.Opazo, L.Garagallo, D.Rdaic(1991) Polymer Bulletin 27:205
10. A.Opazo, L.Garagallo, D.Rdaic(1994) Makromol.Chem., Macromol.Symp.84:289
11. M.Urzua, A.Opazo, L.Gargallo, D.Radic (1998) Polymer Bulletin 40:63
12. J.F.Masson and R.S.Manley (1991) Macromolecules 24:6670
13. C.Y.Liang and R.H.Marchessault (1959) J.Polym.Sci., part I 37:385, (1960)Part II J.Polym.Sci., 43:71